

# Caffeine and Methyliberine: A Human Pharmacokinetic Interaction Study

Original Research

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## Abstract

**Introduction:** Methyliberine and theacrine are methylurates found in the leaves of various *Coffea* species and *Camellia assamica* var. *kucha*, respectively. We previously demonstrated that the methylxanthine caffeine increased theacrine's oral bioavailability in humans.

**Methods:** Consequently, we conducted a double-blind, placebo-controlled pharmacokinetic study in humans administered methyliberine, theacrine, and caffeine to determine methyliberine's pharmacokinetic interaction potential with either caffeine or theacrine. Subjects received an oral dose of either methyliberine, caffeine, methyliberine plus caffeine, or methyliberine plus theacrine using a randomized, double-blind, crossover design. Blood samples were analyzed using UPLC-MS/MS.

**Results:** Methyliberine exhibited linear pharmacokinetics that were unaffected by co-administration of either caffeine or theacrine. However, methyliberine co-administration resulted in decreased oral clearance ( $41.9 \pm 19.5$  vs.  $17.1 \pm 7.80$  L/hr) and increased half-life ( $7.2 \pm 5.6$  versus  $15 \pm 5.8$  hrs) of caffeine. Methyliberine had no impact on caffeine's maximum concentration ( $440 \pm 140$  vs.  $458 \pm 93.5$  ng/mL) or oral volume of distribution ( $351 \pm 148$  vs.  $316 \pm 76.4$  L).

**Conclusions:** We previously demonstrated theacrine bioavailability was enhanced by caffeine, however, caffeine pharmacokinetics were unaffected by theacrine. Herein, we found that methyliberine altered caffeine pharmacokinetics without a reciprocal interaction, which suggests caffeine may interact uniquely with different methylurates.

**Key Words:** caffeine, methyliberine, theacrine, and pharmacokinetics

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## Introduction

The methylxanthine caffeine is found across the globe in a wide variety of plant genera including *Camellia* (e.g., *C. sinensis*), *Coffea* (e.g., *C. arabica*), *Cola* (e.g., *C. nitida*), *Paullinia* (e.g., *P. cupana*), and *Ilex* (e.g., *I. paraguarensis*)<sup>1</sup>. Young leaves, pericarp, and seeds of *C. liberica* are found to contain the methylurate theacrine (1,3,7,9-tetramethyluric acid)<sup>2</sup>, which confirmed the first description of theacrine in a tea plant<sup>3</sup>. Radioactive caffeine tracer studies designed to explore purine metabolism in the leaves of various *Coffea* species demonstrated that during stage 1 of vegetative development, young plants accumulated caffeine synthesized from theobromine<sup>4</sup>. In stage 2,

caffeine is gradually converted to theacrine, which is then converted in stage 3 to liberine (O(2),1,9-tetramethyluric acid) (stage 3), presumably through the intermediate metabolite methyliberine (O(2),1,7,9-tetramethyluric acid).

Methyliberine has recently been granted new dietary ingredient (NDI) status following completion of a 90-day repeated-dose oral toxicity study<sup>5</sup>. In addition, human adverse event potential studies using methyliberine alone, and in combination with theacrine, found no effect of methyliberine on heart rhythm (electrocardiogram; ECG), resting heart rate, or blood pressure<sup>6</sup>. Anecdotal benefits of methyliberine suggesting reduced onset of action of EMF activity without anxiety has led to methyliberine being “stacked” (i.e., combined) with caffeine and/or theacrine. Because we previously demonstrated the interaction potential between the methylxanthine caffeine and the methylurate theacrine, we hypothesized that caffeine would interact with the methylurate methyliberine. Therefore, the purpose of this study was to determine methyliberine pharmacokinetics and its pharmacokinetic interaction potential with theacrine and caffeine following oral administration to humans.

### Scientific Methods

The study protocol and informed consent were approved by the University of Memphis Institutional Review Board (Proposal # FY2018-490). Study participants were informed of all procedures, potential benefits, and risks associated with the study and provided informed consent prior to any study related procedures. The clinical study was conducted at the University of Memphis in accordance with the US Code of Federal Regulations (CFR) governing Protection of Human Subjects (21 CFR Part 50), Financial Disclosure by Clinical Investigator (21 CFR Part 54), and Institutional Review Board (IRB) (21 CFR Part 56). Moreover, the study adhered to the 1996 guidelines of the International Conference on Harmonization (Good Clinical Practice (GCP)), which is consistent with the Declaration of Helsinki as adopted in 2008.

#### Study Design

Study description and eligibility were previously described<sup>6</sup>. In brief, this was a randomized, double-blind, crossover study designed to assess the pharmacokinetic interaction potential of methyliberine, caffeine, and theacrine in healthy subjects. Subjects were randomized in double blind manner to receive a single oral dose of either methyliberine 25 mg (Cohort I), methyliberine 100 mg (Cohort II), caffeine 150 mg, methyliberine 100 mg and caffeine 150 mg (Cohort III) or methyliberine 100 mg and theacrine 50 mg (Cohort IV). Methyliberine (Dynamine®) and theacrine (TeaCrine®) were provided by Compound Solutions (Carlsbad, CA). Caffeine, administered as caffeine anhydrous, was obtained from Nutravative Ingredients (Allen, TX). Serial blood samples were collected at baseline (pre-dose) and 0.25, 0.5, 1, 1.5, 2, 4, 6, 8, and 24 hours post-dose.

Eight hours post-dose, participants stayed in a conference room and were allowed to study, watch TV, or browse the internet. Meal replacement bars and shakes were provided 2- and 6-hours post-ingestion. Water intake was matched across conditions. Participants then went home with standardized food consisting of meal replacement bars and shakes that was kept consistent between visits. Participants returned the following day 24 hours after supplement ingestion. Participants were expected to arrive in a 10-hour fasted state at the same time for each visit between 6am-8am. Participants were asked to refrain from exercise the day prior to their lab visit. Upon arrival in the lab, participants rested for 10-15 minutes before blood pressure readings and catheter insertion. After the baseline sample was drawn, participants ingested the supplement, *i.e.* time of ingestion was matched across conditions. During screening, participants completed a publicly available caffeine consumption questionnaire. Participants were excluded for consuming greater than 400 mg daily. Participants kept diet logs 2 days prior to each lab visit. Meal replacement bars and shakes, as well as water intake, was matched across conditions

#### LC-MS/MS

Plasma levels of caffeine, methyliberine, and theacrine were measured using a previously described UPLC-MS/MS method<sup>7</sup>. Briefly, bioanalytical method inter- and intra-day accuracy and precision were verified to be  $\pm 15\%$ . The lower limit of quantification for caffeine, methyliberine, and theacrine was 0.67 ng/mL. Plasma samples were extracted with methanol containing the internal standard (caffeine-<sup>13</sup>C<sub>3</sub>). The LC-MS/MS system comprised a Waters Acquity UPLC™ I-class system (Waters Corporation, Milford, MA, USA) coupled with a Xevo TQ-S triple quadrupole mass spectrometry detector operating in the positive electrospray ionization (ESI) mode (capillary voltage, 1.1 kV; source temperature, 150 °C; desolvation temperature, 500 °C; desolvation gas flow, 1000 L/h, and cone gas flow, 150 L/h). Separation was achieved using an UPLC BEH C<sub>18</sub> column (50 mm × 2.1 mm I.D., 1.8 μm) and a mobile phase comprising water containing 0.1 % formic acid (A) and acetonitrile with 0.1 % formic acid (B). Detection was obtained using the Multiple Reaction Monitoring (MRM) mode including two MRMs for confirmation of each analyte. The

quantification MRMs for caffeine, caffeine- $^{13}\text{C}_3$  (IS), theacrine, and methylxanthine were  $m/z$  195.11→138.01, 198.1→140.07, 225.12→168.02, and 225.12→167.95, respectively.

#### Pharmacokinetic Data Analysis

Caffeine, methylxanthine, and theacrine oral pharmacokinetic parameters were estimated from plasma concentration-time data, adjusted for lag time ( $t_{lag}$ ), using noncompartmental methods in Phoenix WinNonlin (version 8.2, Certara USA, Inc., Princeton, NJ) as previously described<sup>8</sup>. Maximum concentration ( $C_{max}$ ) and time corresponding to  $C_{max}$  ( $T_{max}$ ) were determined from the plasma concentration versus time data. Area under the plasma concentration-time curve from time 0 to infinity ( $AUC_{0-\infty}$ ) was calculated using the linear trapezoidal rule. The terminal half-life ( $t_{1/2}$ ), was evaluated using  $\ln 2/k_{el}$ , with  $k_{el}$  as the terminal rate elimination constant estimated from the slope of the linear regression of the log plasma concentration versus time curve during the terminal phase. The oral clearance (CL/F) was calculated by dividing the administered oral dose by  $AUC_{0-\infty}$ . The apparent oral volume of distribution during the terminal elimination phase ( $V_z/F$ ) was calculated by dividing CL/F by  $k_{el}$ .

#### Statistical Analysis

To determine the probability of interaction magnitude between methylxanthine and caffeine and/or theacrine, pharmacokinetic parameters were first logarithmically transformed. For each parameter, mean differences of the transformed values were obtained by taking the average of the difference of the transformed values, and upper and lower confidence level with a 90% confidence interval (CI) were obtained using the paired  $t$  test function in Excel. The results of this analysis were exponentiated which corresponded to 90% confidence intervals around the geometric mean ratios for any observed pharmacokinetic parameters<sup>9</sup>.

## Results

#### Subject characteristics

Twelve healthy men ( $n=6$ ; aged  $24\pm 4$  years;  $77\pm 6$  kg) and women ( $n=6$ ; aged  $22\pm 3$  years;  $58\pm 4$  kg) participated in this study. Men and women ingested a daily amount of caffeine  $155\pm 85$  mg and  $230\pm 103$  mg, respectively. All subjects were well tolerated all treatments and no adverse effect was recorded. Diet intake was not changed across all treatment conditions for total kilocalorie, macro- and micro-nutrient composition.

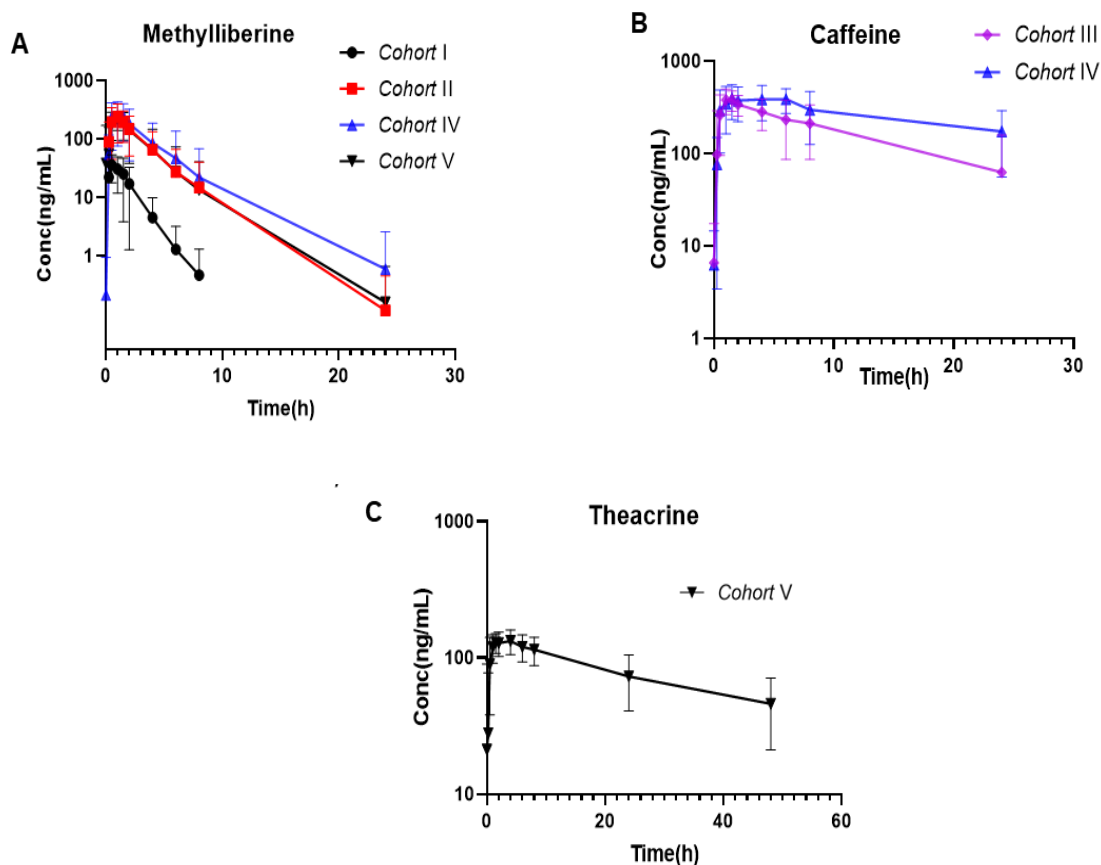
#### Pharmacokinetics

Mean plasma concentration ( $\pm$  standard deviation) time profiles for methylxanthine, caffeine, and theacrine are shown in **Figures 1A-C**. Methylxanthine pharmacokinetic parameters for each cohort are shown in **Table 1**. We found the methylxanthine was rapidly absorbed from the oral administration, with  $C_{max}$  reached on average at 0.6 and 0.9 hours after following low (25 mg) and high (100 mg) doses of methylxanthine, respectively (**Figure 1A, Table 1**). Thereafter, methylxanthine was eliminated with a half-life averaging 1 to 1.4 hours. Dose-normalized  $C_{max}$  and AUC were significantly higher, oral clearance and oral volume of distribution were significantly lower, following 100 mg dose of methylxanthine compared to 25 mg of methylxanthine.

**Table 1.** Methylxanthine pharmacokinetic parameters

Parameter	Cohort I	Cohort II	Cohort IV	Cohort V
$C_{max}$ (ng/mL)	$55.3\pm 34.6$	$287\pm 141$	$349\pm 130$	$289\pm 91.6$
$T_{max}$ (hours)	$0.6\pm 0.3$	$0.9\pm 0.3$	$0.9\pm 0.4$	$0.8\pm 0.5$
$t_{1/2}$ (hours)	$1.0\pm 0.3$	$1.4\pm 0.6$	$1.5\pm 0.8$	$1.4\pm 0.6$
AUC (h x ng/mL/mg)	$3.4\pm 2.2$	$8.0\pm 6.8$	$11.2\pm 11.6$	$7.6\pm 6.6$
CL/F (L/h)	$426\pm 262$	$201\pm 122$	$149\pm 88.9$	$189.3\pm 87.6$
$V_d/F$ (L)	$556\pm 254$	$356\pm 164$	$270\pm 126$	$316\pm 99.6$
MRT (hours)	$1.6\pm 0.5$	$2.4\pm 1$	$2.6\pm 1.2$	$2.3\pm 1.1$

Data are Means  $\pm$  SD



**Figure 1.** Plasma concentrations-time profile of (A) methyllicberine in Cohort I, cohort II, cohort IV, and cohort V; (B) caffeine in cohort III, and cohort IV; and (C) theacrine in cohort V. Data represented as the mean  $\pm$  SD. Cohort I, methyllicberine 25 mg; cohort II, methyllicberine 100 mg; cohort III, caffeine 150 mg; cohort IV, methyllicberine 100 mg and caffeine 150 mg; cohort V, methyllicberine 100 mg and theacrine 50 mg; SD, standard deviation.

After a single dose of methyllicberine 25 mg, the  $C_{max}$  was  $55.3 \pm 34.6$  ng/mL,  $t_{1/2}$  was  $1.0 \pm 0.3$  h, AUC was  $3.4 \pm 2.2$  ng·h/mL/mg, CL/F was  $426 \pm 262$  L/h,  $V_d/F$  was  $556 \pm 254$  L. Compared with methyllicberine 25 mg, oral administration of methyllicberine 100 mg resulted in a decrease in the  $V_d/F$  and CL/F **Table 1**. Based on the geometric means of  $C_{max}$ ,  $T_{max}$ , AUC, CL/F, and  $V_d/F$ , exposure of methyllicberine (100 mg) was different than methyllicberine and when co-administered with caffeine **Table 2**, but was comparable between methyllicberine and when co-administered with theacrine. The geometric ratios for  $C_{max}$ ,  $T_{max}$ , half-life, AUC, CL/F,  $V_d/F$ , and MRT on oral coadministration of methyllicberine (100 mg) with caffeine (150 mg) versus methyllicberine (100 mg) alone were 0.9, 1.24, 1.0, 1.23, 0.81, 0.81, and 1.03 respectively **Table 2**. The geometric ratios for  $C_{max}$ ,  $T_{max}$ , half-life, AUC, CL/F,  $V_d/F$ , and MRT on oral coadministration of methyllicberine (100 mg) with theacrine (50 mg) versus methyllicberine (100 mg) alone were 1.08, 1.03, 0.95, 0.95, 1.05, 0.99, and 1.03 respectively.

**Table 2.** Summary of the statistical analysis of the pharmacokinetic parameters of methyllicberine after single oral administration of a 100 mg dose of methyllicberine alone and in co-administration with caffeine (150 mg)

Pharmacokinetic Parameters of Methyllicberine	Geometric Mean		90% CI	
	Methyllicberine (100 mg) (Cohort II)	Methyllicberine (100 mg) + Caffeine (150 mg) (Cohort IV)	mean ratio	
$C_{max}$ (ng/mL)	254	229	0.9	(0.53-1.54)
$T_{max}$ (hours)	0.77	0.96	1.24	(0.84-1.85)
$t_{1/2}$ (hours)	1.40	1.41	1.00	(0.92-1.10)
AUC (h x ng/mL/mg)	6.69	8.26	1.23	(1.04-1.47)
CL/F (L/h)	149	121	0.81	(0.68-0.97)
$V_d/F$ (L)	303	246	0.81	(0.69-0.96)
MRT (hours)	2.31	2.39	1.03	(0.93-1.15)

We found caffeine  $C_{max}$ , and  $T_{max}$  were unaffected by methyllicberine coadministration. However, methyllicberine coadministration significantly increased  $t_{1/2}$  ( $14.7 \pm 5.8$  vs  $7.15 \pm 5.59$  h), and AUC ( $70.8 \pm 36.9$  vs  $30.5 \pm 17.8$  ng·h/mL/mg). Moreover, methyllicberine decreased caffeine oral clearance (CL/F,  $17.1 \pm 7.8$  vs  $41.9 \pm 19.5$  L/h).

After a single dose of caffeine 25 mg, the geometric mean  $t_{1/2}$  was 5.3 h, AUC was 26.78 ng·h/mL/mg, and CL/F was 37.34 L/h. Compared with caffeine 100 mg, oral coadministration of methyllicberine 100 mg with caffeine 150 mg resulted in an increase in the geometric mean  $t_{1/2}$  was 13.6 h, AUC was 63.98 ng·h/mL/mg, and in a decrease in the geometric mean CL/F was 15.63 L/h. The geometric mean ratios for  $C_{max}$ ,  $T_{max}$ , half-life, AUC, CL/F,  $V_d/F$ , and MRT on oral administration of caffeine (150 mg) versus caffeine 150 mg plus methyllicberine 100 mg were 1.04, 1.63, 2.56, 2.39, 0.42, 1.07 and 2.55 respectively.

The study was not designed to determine the effect of caffeine and/or methyllicberine co-administration on theacrine pharmacokinetics, viz., there was not an arm where subjects received only theacrine. However, based on our previous pharmacokinetic studies with theacrine and caffeine, it appears that methyllicberine increased the half-life of theacrine by approximately two-fold<sup>8</sup>.

## Discussion

Variation in caffeine sensitivity has spurred the discovery of wider therapeutic index natural stimulant platforms that include a unique class of purine alkaloids known as methylurates, e.g., theacrine, also exert their pharmacologic effects via adenosine receptor modulation<sup>2,4,10,11,12,13</sup>. Intriguingly, however, the pharmacologic profile of theacrine appears distinct from caffeine in that it does not alter cardiovascular parameters (e.g., heart rate)<sup>14-16</sup>. For this reason, theacrine is frequently combined (“stacked”) with caffeine in energy, mood, and focus dietary supplements; unfortunately, with little regard for pharmacokinetic and pharmacodynamic interaction potential. We previously demonstrated that when combined, caffeine diminished theacrine’s oral clearance (CL/F) without altering its half-life ( $t_{1/2} \sim V_d/CL$ ), which suggested that the most likely mechanism for the observed interaction was that caffeine increased theacrine’s oral bioavailability (F)<sup>8</sup>. In the present study, we expanded our investigation of the interaction potential between caffeine and methylurates by examining the impact of methyllicberine on caffeine pharmacokinetics. Similar to our previous study, we found that caffeine co-administration led to a modest, but significant, decrease in CL/F and  $V_d/F$ , as well as an increase in plasma area under the curve ( $AUC = (F \cdot Dose)/CL$ ) of methyllicberine. However, methyllicberine half-life, and by extension  $V_d$  and CL, were unaltered by caffeine.

In our previous study investigating the pharmacokinetic interaction potential between theacrine and caffeine, theacrine was found to have essentially no effect on caffeine bioavailability or clearance<sup>8</sup>. The inability of theacrine to increase caffeine bioavailability is not surprising as caffeine is a low extraction drug with an oral bioavailability approaching unity. However, the lack of an effect of theacrine on caffeine clearance is informative since it implied that theacrine, while it may be a CYP1A2 substrate, is not a clinically significant CYP1A2 inhibitor. In the current study, however, concomitant administration of caffeine and methyllicberine led to significantly increased caffeine exposure (AUC), which was accompanied by commensurate decreases in half-life and oral clearance (CL/F). Interestingly, data from our previous study, although not designed to evaluate pharmacokinetic interaction potential, clearly showed that caffeine oral clearance (CL/F) was substantially lower than literature reports when co-administered as a cocktail also

containing both theacrine and methylxanthine<sup>7</sup>. The mechanism by which methylxanthine reduced oral clearance (CL/F) of caffeine is unlikely related to increased bioavailability (F) considering the fact that caffeine's bioavailability is complete and that caffeine's oral volume of distribution (Vd/F) was unchanged.

A clue as to the potential mechanism by which methylxanthine decreases the oral clearance (CL/F) of caffeine is provided by the fact that caffeine is a low hepatic extraction drug that is extensively metabolized (>90%) by CYP1A2 to the N3-demethylated metabolite paraxanthine<sup>17</sup>. Hepatic clearance of low extraction drugs is approximated by multiplying the fraction of unbound drug ( $f_{up}$ ) and intrinsic clearance ( $CL_{int}$ )<sup>18</sup>. Thus, a reduction in caffeine's hepatic clearance is likely attributable to a reduction in intrinsic clearance, which reflects CYP1A2 activity. Thus, our data support the notion that methylxanthine decreases the intrinsic clearance of caffeine through mechanisms likely including inhibition of CYP1A2. However, we cannot discount many other potential factors such as gender, race, genetic variation, disease, and exposure to inducers, which contribute to large interindividual variability in CYP1A2 activity and thus caffeine clearance<sup>17,19,20</sup>. For example, caffeine's plasma clearance is reduced in patients with liver cirrhosis, hepatitis B, and hepatitis C<sup>21,22</sup>. Moreover, smoking stimulates caffeine clearance via CYP1A2 induction, whereas cessation of smoking decreases caffeine clearance<sup>20,23-25</sup>. It is also puzzling that theacrine, which was administered at doses similar to methylxanthine doses in this study, did not affect caffeine clearance in our previous study<sup>8</sup>.

### Conclusions

In conclusion, methylxanthine, a methylurate analog of caffeine, increased plasma exposure and half-life of caffeine following concomitant oral administration. The mechanism underlying this pharmacokinetic interaction is likely attributable to methylxanthine inhibition of CYP1A2, which is a major determinant of intrinsic clearance, and thus hepatic clearance, of caffeine. Several important consequences, with regard to herb drug interaction potential, can be inferred from the data assuming reproducibility in larger more diverse populations. First, caffeine is commonly used as a probe drug to examine CYP1A2-mediated drug interactions<sup>26</sup>. Consequently, our data demonstrate that methylxanthine has the potential to interact with other drugs whose elimination depends on CYP1A2. Secondly, methylurate pharmacology is still in its infancy, but early studies imply that methylxanthine and methylxanthine ligands differ in their affinity and selectivity for the adenosine A<sub>1</sub> and A<sub>2A</sub> receptors<sup>12</sup>, as well as, the sirtuin 3 receptor<sup>27</sup>. Ergo, additional pharmacology studies are needed to provide insight into the pharmacodynamic interaction potential between methylxanthines and methylurates.

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### References

1. Smit HJ, Rogers PJ. Effects of low doses of caffeine on cognitive performance, mood and thirst in low and higher caffeine consumers. *Psychopharmacology (Berl)*. 2000;152(2):167-173.
2. Wanner H, Peřáková M, Baumann TW, et al. O (2), 1, 9-Trimethyluric acid and 1, 3, 7, 9-tetramethyluric acid in leaves of different Coffea species. *Phytochemistry*. 1975;14(3):747-750.
3. Johnson TB. Purines in the Plant Kingdom: The Discovery of a New Purine in Tea1. *Journal of the American Chemical Society*. 1937;59(7):1261-1264.
4. Petermann JB, Baumann TW. Metabolic Relations between Methylxanthines and Methyluric Acids in Coffea L. *Plant Physiol*. 1983;73(4):961-964.
5. Murbach TS, Glavits R, Endres JR, et al. A Toxicological Evaluation of Methylxanthine (Dynamine(R)). *J Toxicol*. 2019;2019:4981420.
6. Bloomer R, Butawan M, Pence J. Acute impact of a single dose of Dynamine®, TeaCrine®, caffeine, and their combination on systemic hemodynamics and associated measures in men and women. *Medical Research Archives*. 2020;8(4).
7. Wang YH, Mondal G, Butawan M, Bloomer RJ, Yates CR. Development of a liquid chromatography-tandem mass spectrometry (LC-MS/MS) method for characterizing caffeine, methylxanthine, and theacrine pharmacokinetics in humans. *J Chromatogr B Analyt Technol Biomed Life Sci*. 2020;1155:122278.
8. He H, Ma D, Crone LB, et al. Assessment of the Drug-Drug Interaction Potential Between Theacrine and Caffeine in Humans. *J Caffeine Res*. 2017;7(3):95-102.
9. Food and Drug Administration. Guidance for Industry: Drug Interaction Studies —Study Design, Data Analysis, Implications for Dosing, and Labeling Recommendations. US Department of Health and Human Services (2012). Available from:

- <http://www.fda.gov/downloads/drugs/guidancecomplianceregulatoryinformation/guidances/ucm292362.pdf>. In.
10. Petermann J, Baumann TW, Wanner H. A new tetramethyluric acid from *Coffea leberica* and *C. dewevrei*. *Phytochemistry*. 1977;16(5):620-621.
  11. Baumann T, Wanner H. The 1, 3, 7, 9-tetramethyluric acid content of cupu (*Theobroma grandiflorum* Schum.). *Acta Amazonica*. 1980;10(2):425-425.
  12. Feduccia AA, Wang Y, Simms JA, et al. Locomotor activation by theacrine, a purine alkaloid structurally similar to caffeine: involvement of adenosine and dopamine receptors. *Pharmacol Biochem Behav*. 2012;102(2):241-248.
  13. Benowitz NL. Clinical pharmacology of caffeine. *Annu Rev Med*. 1990;41:277-288.
  14. Kuhman DJ, Joyner KJ, Bloomer RJ. Cognitive Performance and Mood Following Ingestion of a Theacrine-Containing Dietary Supplement, Caffeine, or Placebo by Young Men and Women. *Nutrients*. 2015;7(11):9618-9632.
  15. Taylor L, Mumford P, Roberts M, et al. Safety of TeaCrine®, a non-habituating, naturally-occurring purine alkaloid over eight weeks of continuous use. *Journal of the International Society of Sports Nutrition*. 2016;13:2.
  16. Ziegenfuss TN, Habowski SM, Sandrock JE, Kedia AW, Kerksick CM, Lopez HL. A Two-Part Approach to Examine the Effects of Theacrine (TeaCrine®) Supplementation on Oxygen Consumption, Hemodynamic Responses, and Subjective Measures of Cognitive and Psychometric Parameters. *Journal of dietary supplements*. 2016:1-15.
  17. Arnaud MJ. Pharmacokinetics and metabolism of natural methylxanthines in animal and man. *Handb Exp Pharmacol*. 2011(200):33-91.
  18. Wilkinson GR, Shand DG. A physiological approach to hepatic drug clearance. *Clinical Pharmacology & Therapeutics*. 1975;18(4):377-390.
  19. Yang A, Palmer AA, de Wit H. Genetics of caffeine consumption and responses to caffeine. *Psychopharmacology (Berl)*. 2010;211(3):245-257.
  20. Nehlig A. Interindividual Differences in Caffeine Metabolism and Factors Driving Caffeine Consumption. *Pharmacol Rev*. 2018;70(2):384-411.
  21. Scott NR, Stambuk D, Chakraborty J, Marks V, Morgan MY. Caffeine clearance and biotransformation in patients with chronic liver disease. *Clin Sci (Lond)*. 1988;74(4):377-384.
  22. Desmond PV, Patwardhan RV, Johnson RF, Schenker S. Impaired elimination of caffeine in cirrhosis. *Dig Dis Sci*. 1980;25(3):193-197.
  23. Kalow W, Tang BK. Caffeine as a metabolic probe: exploration of the enzyme-inducing effect of cigarette smoking. *Clin Pharmacol Ther*. 1991;49(1):44-48.
  24. Parsons WD, Neims AH. Effect of smoking on caffeine clearance. *Clin Pharmacol Ther*. 1978;24(1):40-45.
  25. Brown CR, Jacob P, 3rd, Wilson M, Benowitz NL. Changes in rate and pattern of caffeine metabolism after cigarette abstinence. *Clin Pharmacol Ther*. 1988;43(5):488-491.
  26. Perera V, Gross A, McLachlan A. Measurement of CYP1A2 Activity: A Focus on Caffeine as a Probe. *Current drug metabolism*. 2012;13:667-678.
  27. Wang GE, Li YF, Zhai YJ, et al. Theacrine protects against nonalcoholic fatty liver disease by regulating acylcarnitine metabolism. *Metabolism*. 2018;85:227-239.